

## Supplementary Materials for

### **Plakophilin-2 truncating variants impair cardiac contractility by disrupting sarcomere stability and organization**

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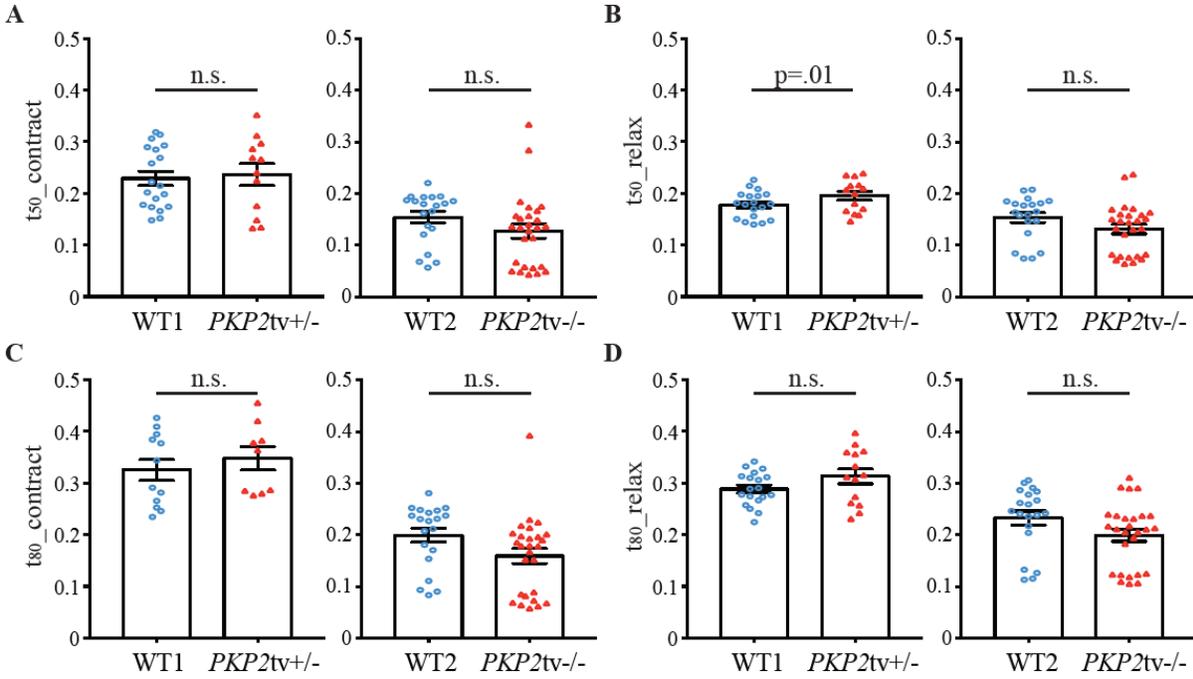
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#### **The PDF file includes:**

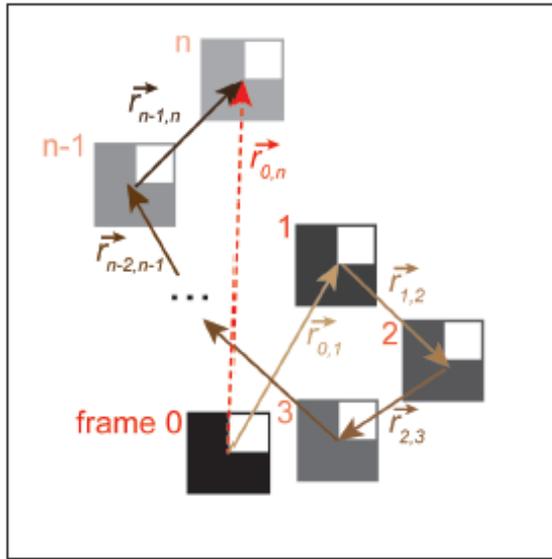
Figs. S1 to S4  
Legends for movies S1 to S12

#### **Other Supplementary Material for this manuscript includes the following:**

Movies S1 to S12



**Fig. S1. Contraction and Relaxation Kinetics of Cardiac Micro-Tissues Electrically Paced at 1Hz. Related to Figure 4.** Quantification of (A) t50\_contract, (B) t50\_relax, (C) t80\_contract and (D) t80\_relax, shows comparable contraction and relaxation kinetics of cardiac micro-tissues made with *PKP2*tv+/- (n = 22) vs. WT1 (n = 25) and *PKP2*tv-/- (n = 27) vs. WT2 (n = 20), when the tissues are electrically paced at 1Hz. **Statistics:** individual data points across three independent experiments are shown with mean  $\pm$  s.e.m. on plots; Student's t-tests,  $\alpha = 0.05$ .



Net Displacement ( $\mu\text{m}$ )

$$\vec{r}_{0,n} = \sum \vec{r}_{i,i+1}$$

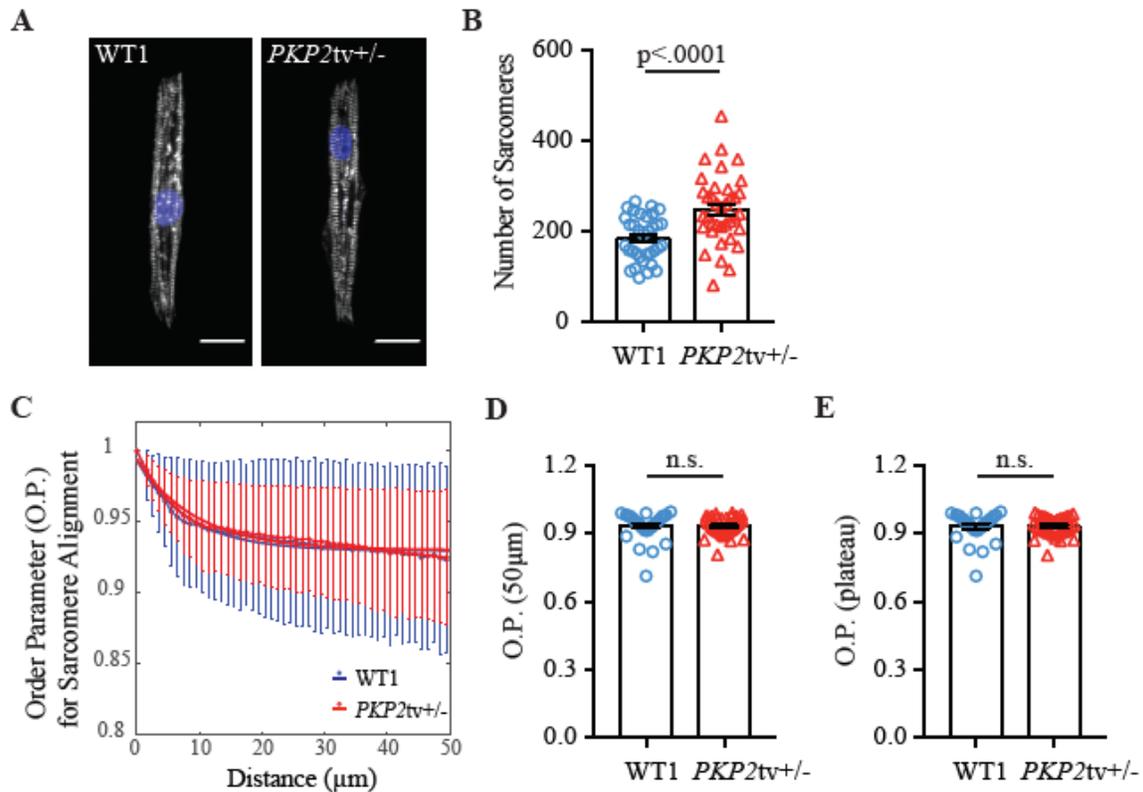
Mean Displacement ( $\mu\text{m}/\text{frame}$ )

$$\frac{1}{n} \sum |\vec{r}_{i,i+1}|$$

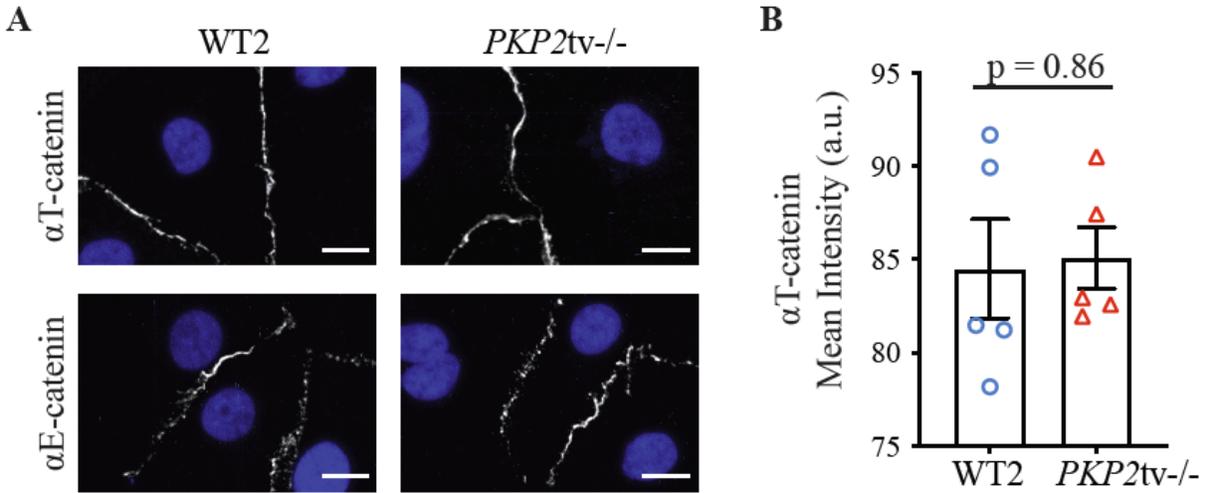
Ramble ( $\mu\text{m}/\text{frame}$ )

$$\frac{1}{n} \left[ \sum |\vec{r}_{i,i+1}| - |\vec{r}_{0,n}| \right]$$

**Fig. S2. Diagram of Optical Flow Tracking Algorithm Method and Outputs. Related to Figure 8 and Figure 9.**



**Fig. S3. *PKP2tv* Lead to Increase in Sarcomere Content in Single Cardiomyocytes. Related to Figure 10.** (A) Representative images of single iPSC-CMs micropatterned in rectangular shape, immune-stained for sarcomeric  $\alpha$ -actinin-2 (shown in grey) and nuclei (shown in blue). Scale bars, 20  $\mu\text{m}$ . (B) Quantification of the number of  $\alpha$ -actinin-2-positive sarcomeric Z-discs in single cardiomyocytes shows significantly higher sarcomere content in *PKP2tv+/-* ( $n = 20$ ) vs. WT1 ( $n = 21$ ). (C) Order parameter (O.P.) for sarcomere alignment plotted over distance from any sarcomere Z-disc comparing single cardiomyocytes harboring *PKP2tv+/-* ( $n = 20$ ) vs. WT1 ( $n = 21$ ). Data shown as mean  $\pm$  standard deviation, with the solid lines showing data fitted in an exponential decay model. (D) Quantification of O.P. at 50  $\mu\text{m}$  away from any sarcomere Z-disc shows comparable sarcomere alignment in single cardiomyocytes harboring *PKP2tv+/-* ( $n = 20$ ) vs. WT1 ( $n = 21$ ). (E) Quantification of plateau of O.P. decay model shows significantly comparable sarcomere alignment in long range in single cardiomyocytes harboring *PKP2tv+/-* ( $n = 20$ ) vs. WT1 ( $n = 21$ ). **Statistics:** individual data points across three independent experiments are shown with mean  $\pm$  s.e.m. on plots, unless otherwise specified; Student's t-tests,  $\alpha = 0.05$ . Immunostains are representative of at least three independent experiments.



**Fig. S4. Junctional expression of  $\alpha$ T-catenin and  $\alpha$ E-catenin in *PKP2tv* and WT cardiomyocytes. (A)** Representative images of iPSC-CM monolayers fixed and stained for nuclei (blue),  $\alpha$ T-catenin (grey, top) and  $\alpha$ E-catenin (grey, bottom). Scale bars, 10 $\mu$ m. **(I)** Mean fluorescence intensity of junctional  $\alpha$ T-catenin is comparable in *PKP2tv*<sup>-/-</sup> (n = 5) vs. WT2 (n = 5). **Statistics:** individual data points are shown with mean  $\pm$  s.e.m. on plots; Student's t-tests,  $\alpha = 0.05$ .

**Movie S1. Fluorescence Recovery After Photobleaching of Junctional N-cadherin signal in WT1 iPSC-CMs, Related to Figure 7.** Representative FRAP imaging of WT1 cells expressing N-cadherin-mApple cultured as a multicellular cardiac patch. Images were taken before and immediately after 1 minute of photobleaching of a user defined ROI, at a 30-second frame rate for 30 minutes. Scale bar, 5  $\mu\text{m}$ .

**Movie S2. Fluorescence Recovery After Photobleaching of Junctional N-cadherin signal in *PKP2*<sup>tv+/-</sup> iPSC-CMs, Related to Figure 7.** Representative FRAP imaging of *PKP2*<sup>tv+/-</sup> cells expressing N-cadherin-mApple cultured as a multicellular cardiac patch. Images were taken before and immediately after 1 minute of photobleaching of a user defined ROI, at a 30-second frame rate for 30 minutes. Scale bar, 5  $\mu\text{m}$ .

**Movie S3. Fluorescence Recovery After Photobleaching of Junctional N-cadherin signal in WT2 iPSC-CMs, Related to Figure 7.** Representative FRAP imaging of WT2 cells expressing N-cadherin-EGFP cultured as a multicellular cardiac patch. Images were taken before and immediately after 1 minute of photobleaching of a user defined ROI, at a 30-second frame rate for 30 minutes. Scale bar, 5  $\mu\text{m}$ .

**Movie S4. Fluorescence Recovery After Photobleaching of Junctional N-cadherin signal in *PKP2*<sup>tv-/-</sup> iPSC-CMs, Related to Figure 7.** Representative FRAP imaging of *PKP2*<sup>tv-/-</sup> cells expressing N-cadherin-EGFP cultured as a multicellular cardiac patch. Images were taken before and immediately after 1 minute of photobleaching of a user defined ROI, at a 30-second frame rate for 30 minutes. Scale bar, 5  $\mu\text{m}$ .

**Movie S5. Dynamics of N-cadherin Junctions in WT1 iPSC-CMs, Related to Figure 8.** Representative time-lapse imaging of WT1 cells expressing N-cadherin-mApple cultured in a monolayer. Images were taken at a 10-minute frame rate for 8 hours. Scale bar, 10  $\mu\text{m}$ .

**Movie S6. Dynamics of N-cadherin Junctions in *PKP2*<sup>tv+/-</sup> iPSC-CMs, Related to Figure 8.** Representative time-lapse imaging of *PKP2*<sup>tv+/-</sup> cells expressing N-cadherin-mApple cultured in a monolayer. Images were taken at a 10-minute frame rate for 8 hours. Scale bar, 10  $\mu\text{m}$ .

**Movie S7. Dynamics of N-cadherin Junctions in WT2 iPSC-CMs, Related to Figure 8.** Representative time-lapse imaging of WT2 cells expressing N-cadherin-EGFP cultured in a monolayer. Images were taken at a 10-minute frame rate for 8 hours. Scale bar, 10  $\mu\text{m}$ .

**Movie S8. Dynamics of N-cadherin Junctions in *PKP2*<sup>tv-/-</sup> iPSC-CMs, Related to Figure 8.** Representative time-lapse imaging of *PKP2*<sup>tv-/-</sup> expressing N-cadherin-EGFP cultured in a monolayer. Images were taken at a 10-minute frame rate for 8 hours. Scale bar, 10  $\mu\text{m}$ .

**Movie S9. Dynamics of N-cadherin Junctions and Sarcomeric  $\alpha$ -Actinin-2 in WT1 iPSC-CMs, Related to Figure 9.** Representative time-lapse imaging of WT1 cells expressing N-cadherin-mApple (shown in green) and EGFP- $\alpha$ -Actinin-2 (shown in magenta) cultured in a monolayer. Images were taken at a 10-minute frame rate for 8 hours. Scale bar, 10  $\mu\text{m}$ .

**Movie S10. Dynamics of N-cadherin Junctions and Sarcomeric  $\alpha$ -Actinin-2 in *PKP2*<sup>tv+/-</sup> iPSC-CMs, Related to Figure 9.** Representative time-lapse imaging of *PKP2*<sup>tv+/-</sup> cells expressing N-cadherin-mApple (shown in green) and EGFP- $\alpha$ -Actinin-2 (shown in magenta) cultured in a monolayer. Images were taken at a 10-minute frame rate for 8 hours. Scale bar, 10  $\mu\text{m}$ .

**Movie S11. Dynamics of N-cadherin Junctions and Sarcomeric  $\alpha$ -Actinin-2 in WT2 iPSC-CMs, Related to Figure 9.** Representative time-lapse imaging of WT2 cells expressing N-cadherin-EGFP (shown in green) and mApple- $\alpha$ -Actinin-2 (shown in magenta) cultured in a monolayer. Images were taken at a 10-minute frame frame rate for 8 hours. Scale bar, 10  $\mu$ m.

**Movie S12. Dynamics of N-cadherin Junctions and Sarcomeric  $\alpha$ -Actinin-2 in *PKP2*<sup>tv/-</sup> iPSC-CMs, Related to Figure 9.** Representative time-lapse imaging of *PKP2*<sup>tv/-</sup> cells expressing N-cadherin-EGFP (shown in green) and mApple- $\alpha$ -Actinin-2 (shown in magenta) cultured in a monolayer. Images were taken at a 10-minute frame frame rate for 8 hours. Scale bar, 10  $\mu$ m.